



Consensus guidelines for diagnosis, prophylaxis and management of *Pneumocystis jirovecii* pneumonia in patients with haematological and solid malignancies, 2014

L. Cooley,¹ C. Dendle,^{2,3} J. Wolf,^{4,5} B. W. Teh,⁶ S. C. Chen,^{7,8,9} C. Boutlis^{10,11} and K. A. Thursky^{6,12}

¹Department of Microbiology and Infectious Diseases, Royal Hobart Hospital, Hobart, Tasmania, ²Departments of Infectious Diseases and General Medicine, Monash Medical Centre, Monash Health, Clayton, Victoria, ³Southern Clinical School, Faculty of Medicine, Monash University, Melbourne, Victoria, ⁴Department of Infectious Diseases, St. Jude Children's Research Center, Memphis, Tennessee, USA, ⁵University of Tennessee Health Sciences Center, Memphis, Tennessee, USA, ⁶Department of Infectious Diseases and Infection Control, Peter MacCallum Cancer Centre, East Melbourne, Victoria, ⁷Centre for Infectious Diseases and Microbiology Laboratory Services, ICPMR – Pathology West, Westmead, New South Wales, ⁸Department of Infectious Diseases, Westmead Hospital, Westmead, New South Wales, ⁹Sydney Medical School, The University of Sydney, Sydney, New South Wales, ¹⁰Department of Infectious Diseases, Wollongong Hospital, Wollongong, New South Wales, ¹¹Graduate School of Medicine, University of Wollongong, Wollongong, New South Wales, ¹²Victorian Infectious Diseases Service, The Doherty Institute for Infection and Immunity, Parkville, Victoria

Key words

Pneumocystis carinii, *Pneumocystis jirovecii*, haematological malignancy, solid-organ tumour, prophylaxis, treatment.

Correspondence

Karin Thursky, Department of Infectious Diseases, Peter MacCallum Cancer Centre, Locked Bag Number 1 A'Beckett Street, East Melbourne, Vic. 8006, Australia.
Email: karin.thursky@petermac.org

doi:10.1111/imj.12599

Abstract

Pneumocystis jirovecii infection (PJP) is a common cause of pneumonia in patients with cancer-related immunosuppression. There are well-defined patients who are at risk of PJP due to the status of their underlying malignancy, treatment-related immunosuppression and/or concomitant use of corticosteroids. Prophylaxis is highly effective and should be given to all patients at moderate to high risk of PJP. Trimethoprim-sulfamethoxazole is the drug of choice for prophylaxis and treatment, although several alternative agents are available.

Introduction

Pneumocystis jirovecii infection causes pneumonia (PJP; also known as PCP) in patients with immunosuppression due to underlying malignancy, organ transplantation or other conditions. The infection is best studied in those with human immunodeficiency virus (HIV)/acquired immune deficiency syndrome, as there are significant differences in the clinical features of PJP between HIV-infected and non-infected individuals.^{1–5}

Infection in immunosuppressed HIV-negative individuals has a shorter duration of onset and fewer systemic symptoms than in HIV-infected individuals.¹ Bronchoalveolar lavage fluid from immunosuppressed non-HIV-infected patients shows lower concentrations of organisms but higher inflammatory scores.⁶ Clinical

disease is generally more severe in these patients, with increased length of hospital stay and higher rates of intensive care unit (ICU) admission and mechanical ventilation, compared with HIV-infected patients.^{1–4} Significantly higher mortality rates are also observed in non-HIV-infected individuals (34–39%) compared to those with HIV (6–7%).^{1,5,7}

With a focus on HIV-negative populations, this guideline outlines diagnostic approaches and provides recommendations for prophylaxis and treatment of PJP in adults and children undergoing chemotherapy for haematological and solid-organ tumours. It does not discuss *P. jirovecii* affecting extra-pulmonary sites or infection in the setting of organ transplantation.

Methodology

Questions asked

In preparing this update, we aimed to address the following questions:

Conflicts of interest: The following authors are consultants or advisory committee members or receive honoraria, fees for service, or travel assistance from, or have research or other associations with the organisations listed: Sharon Chen – Gilead, Merck Sharp & Dohme, Pfizer; Karin Thursky – Pfizer.

- 1 What are the current diagnostic strategies for PJP?
- 2 Which patients with haematological and solid malignancies are at risk for PJP?
- 3 What is the evidence for prophylaxis? Which agents are most effective?
- 4 What are the first- and second-line agents for treatment of PJP?
- 5 Is there any evidence to keep patients with confirmed PJP in clinical isolation?

Search strategy

A literature review was performed using PubMed to identify papers published since 2007 that pertained to PJP in patients with haematological and solid tumours. Search terms included (in combination) 'Pneumocystis carinii', 'Pneumocystis jirovecii', 'immunocompromised', 'non-HIV', 'treatment', 'prophylaxis' and 'diagnostics'.

Diagnosis of PJP

Microbiological tests

Due to difficulties with isolating and culturing this pathogen, microscopic examination of respiratory specimens using various staining methods is used to visualise and identify the morphological structures of *P. jirovecii*. Methenamine silver and toluidine blue preparations stain only the cyst wall and do not allow detection of trophozoites. However, Giemsa stains detect all life stages of *P. jirovecii*. Direct and indirect immunofluorescent assays (DFA, IFA) are specific for different life stages, depending on the antibody used. Comparative studies have shown DFA and IFA to be the most sensitive stains for *P. jirovecii* in sputum and bronchoalveolar lavage, with sensitivities of 97% and 90% respectively.⁸ These assays are also commercially available.

Overall, the accuracy of staining methods is highly dependent on the quality of respiratory specimen, sample processing, reaction of the specimen to the stain chosen and experience of the laboratory observer. The lower burden of *P. jirovecii* in non-HIV-immunocompromised patients, and the likelihood that they may already be on anti-pneumocystis prophylaxis, remains a challenge for diagnosis.⁹

Staining methods have now largely been supplanted by highly sensitive molecular techniques, using semi- or fully quantitative polymerase chain reaction (PCR) targeting *P. jirovecii*-specific genes.¹⁰ A meta-analysis of PCR studies has shown a pooled sensitivity of 99% and specificity of 92% in the non-HIV patient population.¹¹ A variety of gene targets can be used for PCR amplifi-

cation, including: major surface glycoprotein (*MSG*) gene, mitochondrial large subunit (*mtLSU*) rRNA gene, dihydropteroate synthase (*DHPS*) gene, dihydrofolate reductase (*DHFR*) gene, heat shock protein (*HSP*) 70 gene and the beta-tubulin gene. Multicopy genes such as *mtLSU* and *MSG* offer the greatest sensitivity for *P. jirovecii* detection.¹⁰ However, they lack specificity and have a low positive predictive value due to the increased detection of *P. jirovecii* in patients who are colonised but otherwise well. With a high negative predictive value, PCR is best utilised for excluding the diagnosis of PJP.¹²

Quantitative PCR (qPCR), with defined upper- and lower-quantitation thresholds of *P. jirovecii* copy number, can be used to distinguish true infection from colonisation.^{13–18} However, there remains an indeterminate zone of unclear clinical significance.^{14–17} Further, these quantitation thresholds will vary between different patient populations; clinicians should be cautious of relying on them as a measure to guide diagnosis or therapy. To improve accuracy, combined diagnostic algorithms have been proposed. These use qPCR and $\beta(1,3)$ -D-glucan on serum testing to classify patients into infection and colonisation categories,¹⁹ but validation is still required in patients with malignancy.

Due to heterogeneity of methods and diversity of chosen qPCR targets in published literature, generalisability of reported cut-off values is limited. Each institution must establish and validate its own qPCR cut-off values appropriate to the preferred molecular method. While molecular tests may aid clinical diagnosis, there is a lack of correlation between qPCR and the clinical features or outcomes of PJP.²⁰ Therefore, clinicians should take an individual's clinical risk of PJP into consideration when interpreting the cycle thresholds.

Studies on the use of molecular diagnosis have largely been based on bronchoalveolar lavage sampling.^{15,16} Although induced sputum samples have also been evaluated, they can be demanding to perform on wards and in children.¹⁷ Others have reported the use of oral washes and expectorated sputum, but the clinical validity of these approaches requires further evaluation.^{20,21} These approaches certainly produce a lower yield – that may be useful – but, overall, their value in this particular setting remains unclear.

Radiological findings

Imaging studies are an essential companion to microbiological testing for diagnosing PJP. High-resolution computed tomography (CT) is the most commonly used and reliable radiological modality for detecting pneumonic

changes in immunocompromised patients.²² The most frequent CT findings are bilateral, ground-glass changes with apical predominance and peripheral sparing.²² In non-HIV-immunocompromised patients with PJP, signs of consolidation are detected more frequently on CT – and cystic changes *less* frequently – than in HIV-positive patients with PJP.^{23,24} The range of other radiological features seen in PJP include a combination of ground glass and consolidative opacities, cystic changes, linear-reticular opacities, solitary or multiple nodules and parenchymal cavities.²⁵ With treatment, the vast majority of these changes resolve.²²

Nuclear imaging modalities such as 18F-fluorodeoxyglucose-positron emission tomography (FDG-PET) have been used as an adjunct to plain X-ray or CT.²⁶ The FDG-PET findings correlate well with conventional imaging, showing bilateral uptake of FDG in the upper zones of the lungs.²⁷ Abnormalities on PET scans, leading to PJP diagnosis in the setting of normal chest radiographs, have been reported, which suggests a potential role for PET scans in facilitating early diagnosis. However, limited access to FDG-PET is currently a significant barrier to use.²⁸

Determining patient risk and the need for PJP prophylaxis

Antimicrobial prophylaxis is highly successful in preventing PJP in patients with immunosuppression from a diverse range of causes, including solid-organ transplantation and malignancy. Chemoprophylaxis was first demonstrated to be highly effective in preventing PJP in a randomised controlled study of paediatric oncology patients.²⁹ Pooled data from 11 clinical trials showed a reduction in the relative risk (RR) of PJP (0.09, 95% confidence interval (CI) 0.02–0.32) in patients receiving trimethoprim-sulfamethoxazole (TMP-SMX) prophylaxis compared with patients receiving no intervention, placebo or an antibiotic without PJP activity.³⁰ TMP-SMX prophylaxis has been shown to have a mortality benefit and its use is recommended in settings where the risk of PJP is estimated to exceed 3.5% (grade A recommendation).³⁰ Prophylaxis is routinely used in children for a wider range of malignancies, including those with a lower risk of PJP, as TMP-SMX is very well tolerated.

In patients with haematological malignancy, both host and disease factors, as well as chemotherapy regimens need to be considered when determining an individual's risk of PJP (see Table 1). A patient's risk of PJP is influenced by multiple factors at any given point in time and risk should be continually assessed throughout the treatment period.

Table 1 Patients with malignancy at high risk for *Pneumocystis jirovecii* pneumonia

- Heavily pre-treated patients (e.g. multiple lines of chemotherapy for Hodgkin and non-Hodgkin lymphoma, lymphoproliferative diseases or myeloma).
- Relapsed disease.
- High-dose corticosteroids (often in combination with chemotherapy).
- Chemotherapy, monoclonal antibodies or diseases causing prolonged lymphopenia (e.g. ALL, alemtuzumab, R-CHOP14).

PJP risk in patients with haematological malignancy

Specific patient groups with underlying haematological malignancy have been identified to be at high risk of PJP.³¹ These findings have been used to inform guidelines regarding institution of PJP prophylaxis. However, there have been important shifts in the treatment of haematological malignancies^{30,32} and diagnostic algorithms since the publication of earlier studies. Furthermore, early evaluation did not encompass emerging subgroups of patients with haematological malignancy. The estimated risk of PJP in these groups, such as acute myeloid leukaemia,^{3,33,34} myelodysplasia, myeloma^{3,35–37} and lymphoma,^{38–46} are based on case reports and are incompletely defined.

The most easily quantifiable risk factors in patients with haematological malignancy are choice, dose and combination of chemotherapy agents. Other factors, such as age, comorbidities, type⁴⁷ and stage of malignancy, surgery, radiation and immune parameters,⁴⁵ also contribute to risk but are more difficult to quantify.

Patients at highest risk of PJP are those undergoing treatment for acute lymphoblastic leukaemia (ALL) (up to 16%)⁴⁸ and for allogeneic haemopoietic stem cell transplant (HSCT) (from 0.3% to 15%).^{3,49,50} In these populations, the number needed to treat has been estimated to be 11.³⁰

Patients receiving prolonged, high-dose corticosteroid treatment (16–25 mg of prednisolone per day or ≥ 4 mg dexamethasone daily for ≥ 4 weeks) are also at high risk of PJP, regardless of underlying type or stage of malignancy, or use of other chemotherapy agents.⁵¹ Certain T-cell-depleting agents (e.g. alemtuzumab) will also place patients at a higher risk of PJP, particularly if salvage treatment is being administered.⁵² Further, patients who have had PJP previously remain at high risk of a subsequent episode if their underlying immune deficit persists.

Patients in whom the risk of PJP is not conclusively established, but is likely to be moderate, include those with autologous bone marrow transplant,³ and patients

with haematological malignancy undergoing certain high-intensity chemotherapy regimens. In the latter case, clinicians should be particularly vigilant of R-CHOP14 (rituximab, cyclophosphamide, adriamycin, vincristine, prednisolone chemotherapy on a 14-day cycle),^{38,40,42,43,53} FCR (fludarabine, cyclophosphamide, rituximab),²⁷ AVBD (adriamycin, vincristine, bleomycin, dexamethasone),⁵⁴ gemcitabine⁵⁵ and high-dose methotrexate. Patients with prolonged CD4 lymphopenia, before or after initiation of chemotherapy, are also likely to be at moderate risk of PJP. However, with limited evidence to assess PJP risk in this group, debate still surrounds the use of prophylaxis in this population.^{56,57}

Patients with haematological malignancy at lower risk of PJP include those receiving low-intensity chemotherapy regimens (such as R-CHOP given on a 21-day cycle), provided they do not have other risk factors.^{38,45,46,58}

PJP risk in patients with solid-organ malignancy

PJP has been reported to occur in patients undergoing chemotherapy for a broad range of solid-organ malignancies, including breast, pulmonary, renal, genitourinary and colorectal cancer; central nervous system tumours; rhabdomyosarcoma and melanoma.^{4,7,59–61} Despite this, there are no clinical efficacy data supporting the routine use of PJP prophylaxis in these patient groups.

The inclusion of corticosteroid therapy in the chemotherapy regimen has been consistently reported as a risk factor for developing PJP in adult patients with solid-organ tumours.^{7,61–63} Adult patients who are receiving high-dose corticosteroid therapy (16–25 mg prednisolone or ≥ 4 mg dexamethasone daily for ≥ 4 weeks) appear to be at greatest risk, both during high-dose therapy and the steroid-tapering period.⁵⁹ Patients receiving these kinds of doses are typically undergoing dose-intensive chemotherapy for lung, breast or brain cancer. There is a paucity of data in the paediatrics setting, although a dosage equivalent to ≥ 2 mg/kg per day of prednisone or equivalent to a total of ≥ 20 mg/day for children who weigh more than 10 kg, particularly when given for more than 14 days, is considered to be T-cell immunosuppressive.⁶⁴

Patients with brain tumours are at particular risk of PJP. While the incidence of PJP is low (1.7%) among patients receiving corticosteroids for brain tumours,⁶⁵ cranial irradiation increases the risk of PJP. Over a 12-month period, PJP has a reported incidence of 6.2% in this population, with a median time to onset of 10 weeks.⁶⁶ Temozolomide, an alkylating agent used for the treatment of glioblastoma multiforme, results in significant lymphopenia (< 500 cells/uL) in 24–100% of

patients, depending on density of dosing.^{67,68} In a phase 2 trial, 79% of patients developed grade 3–4 lymphopenia, with two of the first 15 patients (13%) developing PJP.⁶⁹ Subsequent cases have led to a black-box warning recommending that PJP prophylaxis accompany the use of this agent.^{7,59,70}

Collectively, these data suggest that PJP prophylaxis should be considered in solid-tumour patients undergoing chemotherapy regimens containing 16–25 mg prednisolone or ≥ 4 mg dexamethasone daily for ≥ 4 weeks, as well as patients undergoing intensive treatment for cerebral malignancy (grade C recommendation). Prophylaxis should be continued for 6 weeks after the steroid-tapering period. A summary of agents for PJP prophylaxis and their indications for use are provided in Table 2.

Chemoprophylaxis for PJP

TMP-SMX

TMP-SMX is the first-line prophylactic agent for PJP prevention in adults and children, and the only agent demonstrated to be more effective than placebo in prospective randomised trials (grade A recommendation).^{29,30} Further, limited evidence suggests that TMP-SMX is superior to other agents, especially in younger patients (< 2 years of age) and in patients undergoing HSCT (grade D recommendation).⁷¹ The use of TMP-SMX may also confer some protection against other infections in the high-risk patient such as toxoplasmosis, nocardia and bacterial sepsis. A systematic review in the HIV population also found that the use of TMP-SMX confers some protection against the development of resistance to other antibiotics.⁷²

The optimal dose schedule for TMP-SMX is not clear due to the limited number of studies comparing regimens in patients with malignancy or undergoing stem cell transplantation.^{29,30,73} Once- or twice-weekly prophylaxis or 3 non-consecutive days per week may be of equal efficacy, based on retrospective and observational studies (level III-2 evidence).⁷⁴ Hughes *et al.* reported that daily and thrice-weekly dosing in children with ALL was equally efficacious.⁷⁵ Studies in HSCT patients have reported PJP rates of $< 1\%$ using doses as low as two single strength (SS) tablets twice weekly, two double strength (DS) tablets twice weekly and two DS tablets thrice weekly.^{76–78} In the absence of clear evidence to support various dosing regimens, these guidelines support the use of once-daily dosing in adults with SS or DS or thrice-weekly dosing with DS (grade B recommendation). In children, various regimens are acceptable, although three days per week is preferred practice.

Limitations in TMP-SMX prophylaxis include documented hypersensitivity, renal impairment, drug

Table 2 Indications for *Pneumocystis jirovecii* chemoprophylaxis in patients (adults and children) with malignancy and recommended agent**Haematological malignancy (grade of recommendation)**

Note: Patients may have multiple or cumulative risk factors; consider underlying disease, disease status and treatment-related immunosuppression.

Known indications:

- Allogeneic HSCT (all) (C)
- ALL (all) (A)
- AML or lymphoma regardless of treatment protocol (children only) (C)
- Autologous HSCT (all children and selected high-risk adults) (C)
- Regimens: R-CHOP14, high-dose methotrexate (C)
- Lymphocyte-depleting agents (e.g. alemtuzumab) or patients whose CD4 count <200 cells/uL before commencing chemotherapy (C)
- Corticosteroids: where 16–25 mg prednisolone or ≥4 mg dexamethasone for ≥4 weeks is planned (C)
- Regimens: FCR, ABVD, gemcitabine (single centre reports of higher risk for these regimens; consider prophylaxis) (D)

Solid tumours (grade of recommendation)

Regimens where 16–25 mg prednisolone or ≥4 mg dexamethasone for ≥4 weeks is planned (C)

Brain tumours, particularly if temozolomide or craniospinal irradiation is planned (B)

Other solid tumours undergoing myelosuppressive chemotherapy (children only) (C)

First-line prophylactic agent should be trimethoprim-sulfamethoxazole, unless:

- | | |
|------|---|
| (i) | Previous allergy or hypersensitivity to sulfa-drugs
Recommend: trimethoprim-sulfamethoxazole desensitisation (unless previous anaphylaxis) |
| (ii) | Planned methotrexate chemotherapy in adults
Recommend: second-line prophylactic agent |

A second-line prophylactic agent should be used if trimethoprim-sulfamethoxazole is contraindicated:

- | | |
|-------|--------------------------------------|
| (i) | Dapsone, OR |
| (ii) | Pentamidine (nebulised, monthly), OR |
| (iii) | Atovaquone |

Prophylaxis should continue for at least 6 weeks after steroid cessation. A longer period of prophylaxis may be required if ongoing chemotherapy (e.g. cytarabine, cyclophosphamide, fludarabine, fluorouracil, methotrexate) is planned. Life-long prophylaxis should be considered if the patient has had a previous episode of PJP and persisting immunosuppression.

interactions, myelosuppression and gastrointestinal disturbance. The true rate of adverse reactions is unknown, but in the adult HSCT population it is estimated to be in the range of 5–15%, and in children it is much lower.⁷⁷

Although one early study suggested increased duration of neutropenia associated with TMP-SMX,⁷⁹ this study pre-dated current chemotherapy regimens (including use of granulocyte colony-stimulating factor). More contemporary studies have not demonstrated an impact upon degree or duration of neutropenia. It is reasonable to administer TMP-SMX prior to engraftment in children receiving HSCT, as there is no evidence of delayed engraftment in this population (level III-2 evidence).⁸⁰ Myelosuppression may be exacerbated when TMP-SMX is used in combination with methotrexate, although clinical data in adults are conflicting. Concurrent methotrexate administration is not a contraindication for use of TMP-SMX in children receiving treatment for cancer, as there is no evidence for a common, clinically significant adverse drug interaction in this population (level III-2 evidence).⁸¹

TMP-SMX should be used with caution in all patients with renal and hepatic impairment, and glucose-6-phosphate dehydrogenase deficiency (G6PD).

Mutations in the *fas* gene of *P. jirovecii*, which encodes the DHPS protein, are associated with prior exposure to sulphonamides.^{82,83} When these mutations are present, reduced susceptibility to TMP-SMX should be expected. However, the mutations do not appear to correlate with resistance, as HIV-infected patients with *Pneumocystis* isolates containing these mutations respond to TMP-SMX therapy.^{84,85} Helweg-Larsen *et al.*⁸⁴ reported an association with increased mortality; however, subsequent studies have failed to detect such an effect.^{85,86} At present, no recommendation can be made with respect to clinical indications or interpretation of resistance testing.

Oral desensitisation regimens have been used successfully for HIV-infected patients and others with rash, and similar protocols have been used in HSCT patients with a success rate of approximately 80%.⁷⁷ Desensitisation may be attempted whenever feasible (level II evidence, grade C recommendation).³² Desensitisation may be considered for all patients with TMP-SMX-associated rash but is contraindicated in those with a prior history of associated drug rash with eosinophilia and systemic symptoms ('DRESS'), Stevens-Johnson syndrome, or toxic epidermal necrolysis.

Second-line agents for PJP chemoprophylaxis

There are no sufficiently powered trials on which to base a recommendation for alternative prophylactic regimens in patients with malignancies. Many recommendations have been based on studies involving patients with HIV infection. However, these should be interpreted with caution due to possible differences in efficacy and side-effect profiles in patients with malignancy. An alternative agent should be used if TMP-SMX is withheld for any reason for ≥ 2 weeks (grade D recommendation).

Dapsone

Dapsone is a synthetic sulfone that acts against *P. jirovecii* by inhibiting the synthesis of dihydrofolic acid through competition with para-amino-benzoate for the active site of dihydropteroate synthetase.⁸⁷ It has 70–80% oral bioavailability with oral dosing achieving high levels of concentration in bronchoalveolar lavage fluid.⁸⁸ Adverse reactions to dapsone include agranulocytosis, aplastic anaemia, rash, nausea and sulfone syndrome (rash, fever, hepatitis, lymphadenopathy and methemoglobinemia).⁸⁹ Dapsone should not be given to patients with G6PD deficiency or to patients who have experienced severe side-effects with TMP-SMX.

Adverse effects requiring discontinuation of therapy occur in up to 43% of HSCT patients, the most common being rash and haemolytic anaemia. Dapsone is not myelosuppressive.⁹⁰ Azole antifungal agents elevate dapsone levels so concomitant use should be avoided or monitored with caution.

Dapsone 50 mg BD three times weekly was compared with TMP-SMX DS BD twice weekly in a study of allogeneic HSCT patients intolerant of TMP-SMX.⁷⁷ The incidence of PJP was significantly higher in the dapsone group (7.2% vs 0.37%), with a relative risk of 18.8 (95% CI 4.0–88.6) – although it should be noted that the dose of dapsone used in this study is lower than that currently used in clinical practice (100 mg daily). In a retrospective study of HSCT patients, Vasconcelles *et al.* reported a failure rate of 3%.⁷⁸

Pentamidine

Pentamidine isethionate is an aromatic diamidine derivative available for parenteral or inhalation use. Given the high rates of adverse effects associated with parenteral therapy in adults, including pancreatitis, hypoglycaemia (27%) and nephrotoxicity (25%), aerosolisation is the preferred mode of delivery for prophylaxis. Nebulised pentamidine is usually well tolerated, with the major side-effects being coughing and wheezing, which can be

prevented by the use of inhaled beta-agonists. It should be avoided in patients less than 5 years of age or if there is a history of asthma. In children, intravenous pentamidine is used during the pre-engraftment period in some centres and may be an alternative if no other options are available. However, paediatric data are limited and it should be used with caution if risk factors for pancreatitis exist.^{91,92}

In one study of patients undergoing HSCT, nebulised pentamidine had significantly lower treatment-related toxicity compared with TMP-SMX (RR 0.19, 95% CI 0.04–0.89).⁷⁸ Aerosolised pentamidine has the convenience of administration at a dose of 300 mg monthly or 150 mg fortnightly. It must be administered through jet-nebuliser in a single or negative-pressure room (grade C recommendation).

Few clinical trials have been undertaken in patients with malignancy. The largest, a retrospective study conducted in patients undergoing HSCT, compared aerosolised pentamidine with TMP-SMX and dapsone. Pentamidine was the least effective, with a higher incidence of PJP: 9.1% compared with 0% in patients treated with TMP-SMX. Significantly, patients who received pentamidine had increased rates of non-PJP infections and a higher mortality 12 months post-transplant compared with patients receiving TMP-SMX or dapsone.⁷⁸

Atovaquone

Atovaquone is a structural analogue of protozoan ubiquinone. It inhibits the binding of ubiquinone to cytochrome *b*, impairing electron-transport mechanisms in *Pneumocystis* mitochondria. It is available as an oral suspension and requires twice-daily dosing. Bioavailability of the suspension is high and is further augmented by administration with a fatty meal. The most common side-effects include rash, nausea, diarrhoea, elevated transaminases and headache, which are usually mild.

Colby *et al.* conducted a prospective randomised trial following adult autologous SCT comparing atovaquone ($n = 20$) with TMP-SMX ($n = 19$) as PJP prophylaxis. No cases of PJP were reported in either group, although the treatment-associated adverse event rate was significantly higher in patients receiving TMP-SMX (40% vs 0%, $P < 0.003$).⁹³

Duration and dosing regimens for PJP chemoprophylaxis

Prophylaxis should continue for a period of time after the immunosuppressive regimen is ceased. In the setting of corticosteroid-containing regimens, prophylaxis should be continued while steroids are being weaned and/or for

Table 3 Dosing schedule for *Pneumocystis jirovecii* chemoprophylaxis in adults and children with malignancy

	Adult	Children
TMP-SMX	160 + 800 mg (one DS tablet) orally, daily Or 80 + 400 mg (one SS tablet) orally, daily Or 160 + 800 mg (one DS tablet) orally, three times a week†	In children >1 month of age: 75 mg/m ² /dose or 2.5 mg/kg/dose (max 160 mg/dose) orally, 12-hourly, 3 times a week†
Dapsone	100 mg orally, daily	In children ≥1 month of age: 2 mg/kg/dose orally, daily OR 4 mg/kg/dose orally, once a week
Pentamidine	300 mg inhaled through nebuliser, every 4 weeks (administered through a jet-nebuliser producing a droplet size of 1–2 microns)	Intravenous (≥2 years): 4 mg/kg/dose (max 300 mg) every 4 weeks Through inhalation (>5 years): 300 mg inhaled through nebuliser, every 4 weeks (administered through a jet-nebuliser producing a droplet size of 1–2 microns)
Atovaquone	1500 mg orally, daily with a high-fat meal	1–3 months: 30 mg/kg/dose orally, daily 4–24 months: 45 mg/kg/dose (max 1500 mg) orally, daily >24 months: 30 mg/kg/dose (max 1500 mg) orally, daily

†Either non-consecutive or consecutive days is acceptable. DS, double strength; SS, single strength.

a period of 6 weeks after cessation. With some chemotherapy regimens (e.g. alemtuzumab⁵² and FCR²⁷), where there are high rates of late-onset PJP, consideration should be given to extended PJP prophylaxis for up to 12 months, particularly in pre-treated patients. CD4⁺ monitoring has been advocated as a method to quantify risk of disease development and to guide duration of prophylaxis in temozolomide and alemtuzumab regimens.^{52,70} However, further studies are required to establish the utility of CD4⁺ monitoring in the non-HIV patient setting. In patients with ongoing immunosuppression (e.g. graft-vs-host disease), prophylaxis should continue indefinitely.

Dosing regimens for PJP prophylaxis are provided in Table 3.

Treatment of PJP

The treatment recommendations for PJP are summarised in Table 4.

TMP-SMX for the treatment of PJP

In view of its proven clinical efficacy, and the cost and availability of both the IV and oral formulations, TMP-SMX is the preferred therapy for mild, moderate and severe disease for all patients with PJP (grade B recommendation).^{95–97} Initial trials in children using a dose of TMP 20 mg/kg/day and SMX 100 mg/kg/day showed an efficacy of 70%, with mortality due to underlying immunosuppressive disease. To date, no agent has been demonstrated to have outcomes superior to TMP-

SMX. Lower doses (i.e. TMP 15 mg/kg/day and SMX 75 mg/kg/day) have been recommended for mild-moderate disease, and as step-down therapy in HIV-infected patients with severe disease following response to high-dose therapy.⁹⁷ Intravenous therapy is recommended for severe disease due to concerns regarding drug absorption in critically unwell patients; for mild and moderate disease, oral or IV therapy may be administered. Based on HIV literature, recommended treatment duration is 21 days (grade C recommendation).

Role of corticosteroids in the treatment of PJP

The use of corticosteroids in HIV-associated PJP was established following randomised trials in the late 1980s.^{98–101} Arterial oxygenation at diagnosis was recognised as the best prognostic indicator of survival.¹⁰² Adjunctive corticosteroid therapy commenced at the time of PJP therapy prevented the early decline in oxygenation that occurs after initiation of PJP therapy in patients with moderate to severe PJP (arterial-alveolar difference >35 mmHg or an arterial oxygen pressure <70 mmHg). This resulted in a reduction in the likelihood of respiratory failure and death. Patients with milder hypoxaemia (arterial oxygen >70 mmHg) may benefit from corticosteroids, but a difference in respiratory failure and death has not been demonstrated.

Studies in non-HIV-infected patients have produced conflicting results. Retrospective studies^{3,103,104} showed no benefit in non-HIV-infected patients with moderate to severe disease. In contrast, Pareja *et al.*¹⁰⁵ found patients who received adjunctive steroids required shorter duration of mechanical ventilation and ICU admission, and

Table 4 Recommended dosing for treatment of *Pneumocystis jirovecii* infection†

	Adult	Child	Comments
TMP-SMX	5 + 25 mg/kg oral or IV, 8-hourly for 21 days In severe disease, increase to 6-hourly initially	>1 month Dose as for adult	<i>First-line therapy for all levels of severity</i> Consider desensitisation except with known history of immediate hypersensitivity or TEN/SJS
Clindamycin plus primaquine	450 mg clindamycin orally, 8-hourly PLUS 15 mg primaquine orally, daily for 21 days In severe disease, increase clindamycin to 900mg IV, 8-hourly initially then dose as above. Also, increase primaquine to 30 mg	10 mg/kg (max 450 mg) clindamycin orally 8-hourly PLUS 0.25 mg/kg primaquine (max 15 mg) orally, daily for 21 days	<i>Second-line therapy for severe disease</i> Test for G6PD deficiency before treatment with primaquine
Pentamidine	In severe disease where TMP-SMX contraindicated: pentamidine 4 mg/kg (max 300 mg) IV, daily for 21 days	In severe disease where TMP-SMX contraindicated: pentamidine 4 mg/kg (max 300 mg) IV, daily for 21 days	<i>Equal second-line therapy for severe disease</i>
Dapsone plus trimethoprim	100 mg dapsone orally, daily PLUS 5 mg/kg trimethoprim orally, 8-hourly for 21 days	2 mg/kg dapsone (max 100 mg) orally, daily PLUS 5 mg/kg trimethoprim orally, 8-hourly for 21 days	<i>Alternative option as first line therapy for mild-moderate disease</i> There is 20% cross-reaction with sulfonamides and dapsone; contraindicated with immediate hypersensitivity or severe reactions. Consider testing for G6PD deficiency prior to use.
Atovaquone	For mild-moderate diseases: 750 mg orally, 12-hourly for 21 days	<3 months or >24 months: 15–20 mg/kg orally, daily; 3–24 months: 22.5 mg/kg orally, 12-hourly for 21 days	<i>Third-line therapy</i>

†Refer to *Therapeutic Guidelines: antibiotic (version 15)* for further details.⁹⁴ SJS, Stevens Johnson Syndrome; TEN, toxic epidermal necrolysis.

reduced supplemental oxygen use. No differences in the rates of intubation or in-hospital mortality were observed.

Although there are insufficient data to make a recommendation regarding the use of adjuvant corticosteroids in patients with PJP and malignancy, it may be appropriate to co-administer corticosteroids, at doses used in HIV-associated PJP, in patients with moderate to severe PCP.

Alternative agents for the treatment of PJP in the setting of TMP-SMX intolerance

Desensitisation should be considered for all patients with TMP-SMX-associated rash but is contraindicated in those with a prior history of associated drug rash with eosinophilia and systemic symptoms ('DRESS'), Stevens-Johnson syndrome or toxic epidermal necrolysis. If TMP-SMX remains contraindicated, alternative agents include atovaquone, clindamycin plus primaquine and dapsone plus trimethoprim, although there is extremely limited evidence for efficacy in children, and limited data for safety in children with cancer (grade D recommendation). Few clinical studies have assessed second-line therapies in patients with malignancy. Therefore, recommendations for second-line therapy are based on data

from clinical studies involving HIV-infected individuals and are in concordance with the current version of *Therapeutic Guidelines: antibiotic, version 15*.⁹⁴

Intravenous pentamidine has been shown to have similar efficacy to TMP-SMX in HIV-infected individuals with PJP.^{106,107} A retrospective review of first- and second-line therapy for HIV-associated PJP¹⁰⁸ found higher rates of unchanged second-line therapy with pentamidine than with TMP-SMX; however, survival rates were significantly lower when compared with TMP-SMX and clindamycin-primaquine. Pentamidine was associated with a higher risk of death (RR 3.3, 95% CI 2.2–5.0). A comparison of aerosolised and reduced-dose intravenous pentamidine therapy in patients with mild to moderate disease found higher rates of early recrudescence and relapse in the aerosolised group.¹⁰⁹

Clindamycin-primaquine was compared with TMP-SMX in a randomised study of mild and moderately severe PJP in HIV-infected individuals.¹¹⁰ In a study limited by sample size, efficacy of clindamycin-primaquine was similar to oral TMP-SMX or dapsone-trimethoprim for treatment of mild-moderate PJP in HIV-infected individuals in terms of dose-limiting toxicity, therapeutic failure or survival.⁹⁵ Notably, serious

haematological toxicity (neutropenia, thrombocytopenia and methemoglobinaemia) was more common in the clindamycin-primaquine arm.

While dapsone has been shown to be an ineffective treatment as a single agent, a combination of oral dapsone plus trimethoprim was effective in patients with HIV-associated PJP.¹¹¹ Major toxicity was seen in only 13% of patients. Dapsone-trimethoprim has been shown to be equivalent to TMP-SMX and clindamycin-primaquine in mild-moderate disease.⁹⁵

Atovaquone (750 mg TDS) has been compared with TMP-SMX in a randomised trial of HIV-infected individuals with mild-moderate PJP.⁹⁶ Non-response rates were significantly higher in the atovaquone arm, although tolerability was greater than with TMP-SMX. Overall successful therapy, defined by clinical response and no adverse events, was equivalent.

Animal studies have demonstrated that echinocandins, which target $\beta(1,3)$ -D-glucan, have activity against *P. jirovecii*;¹¹² however, clinical trials in humans have not been performed. Data regarding efficacy are restricted to case reports, and these have revealed inconsistent responses.^{113–116} Echinocandins cannot be recommended at this time due to their high cost, lack of an oral formulation and the absence of compelling evidence demonstrating their efficacy (grade D recommendation).

Managing PJP treatment failure

Clinical failure is defined as lack of improvement or worsening of respiratory function documented by reduced arterial oxygen saturation after at least 4–8 days of *Pneumocystis* treatment. Early and reversible deterioration in the first 3–5 days is typical, and clinicians should wait at least 4–8 days before changing therapy due to lack of improvement. Although evidence is limited, intravenous pentamidine, clindamycin-primaquine or addition of an echinocandin can be considered for salvage therapy in cases of treatment failure.¹¹⁷

Treatment failure due to drug intolerance appears less common in malignancy-associated PJP than in HIV infection. Relapse has been reported only rarely in non-HIV associated PJP.

PJP infection-prevention measures

A role for isolation precautions to prevent inter-patient transmission of PJP has not been established for patients with malignancy, although there are data that suggest good infection-control measures prevent transmission in

outbreaks of infection among kidney transplant patients.^{118–121} The predominant method of spread for *Pneumocystis* is believed to be person-to-person transmission through exhaled air. Airborne transmission has been demonstrated in animal studies, and air samples from the rooms of patients with PJP have shown the presence of like organisms in air as far as 8 metres from the patient, suggesting that this may be a route of transmission.^{122–124} It is possible that symptomatic patients might be at higher risk of transmission because of apparent higher density of colonisation.^{17,125}

Staff members and other patients may be at risk of becoming colonised from direct or indirect contact with an infected patient.¹²⁶ However, colonisation of healthy humans and previously infected patients is frequent, without placing individuals at risk of subsequent infection. Most cases of PJP occur sporadically, without a known infected contact.^{127,128}

PJP outbreaks in other patient populations (predominantly renal transplantation) have been reported, suggesting person-to-person transmission on the basis of epidemiology and molecular testing.^{119,125,129–131} However, epidemiologically distinct cases with identical molecular types may also occur.¹¹⁸

While some clinicians and institutions recommend that at-risk patients should not share a hospital room with a patient with known PJP, the current evidence is insufficient to mandate isolation in haematology and solid-tumour groups. It is important that patients at risk of PJP are identified and receive prophylaxis.

Conclusion

Patients with haematological and solid-organ tumours are highly susceptible to PJP due to disease- and treatment-related immunosuppression. In this patient population, PJP is associated with high morbidity and mortality. Appropriate prophylaxis for all moderate- to high-risk patients is a key strategy for improving outcomes, given the proven efficacy of this approach. TMP-SMX is the drug of choice for both prophylaxis and treatment, although several alternative agents are available, as outlined here.

Acknowledgements

The authors would like to thank members of ALLG, ASID and ANZCHOG for their review of the draft document, and Dr Candice O'Sullivan from Wellmark Pty Ltd for her assistance in preparing the manuscript for submission.

References

- 1 Nuesch R, Bellini C, Zimmerli W. Pneumocystis carinii pneumonia in human immunodeficiency virus (HIV)-positive and HIV-negative immunocompromised patients. *Clin Infect Dis* 1999; **29**: 1519–23.
- 2 Matsumura Y, Shindo Y, Iinuma Y, Yamamoto M, Shirano M, Matsushima A *et al*. Clinical characteristics of Pneumocystis pneumonia in non-HIV patients and prognostic factors including microbiological genotypes. *BMC Infect Dis* 2011; **11**: 76.
- 3 Pagano L, Fianchi L, Mele L, Girmenia C, Offidani M, Ricci P *et al*. Pneumocystis carinii pneumonia in patients with malignant haematological diseases: 10 years' experience of infection in GIMEMA centres. *Br J Haematol* 2002; **117**: 379–86.
- 4 Roblot F, Imbert S, Godet C, Kauffmann C, Ragot S, Le Moal G *et al*. Risk factors analysis for Pneumocystis jirovecii pneumonia (PCP) in patients with haematological malignancies and pneumonia. *Scand J Infect Dis* 2004; **36**: 848–54.
- 5 Mansharamani NG, Garland R, Delaney D, Koziel H. Management and outcome patterns for adult Pneumocystis carinii pneumonia, 1985 to 1995: comparison of HIV-associated cases to other immunocompromised states. *Chest* 2000; **118**: 704–11.
- 6 Limper AH. Alveolar macrophage and glycoprotein responses to Pneumocystis carinii. *Semin Respir Infect* 1998; **13**: 339–47.
- 7 Sepkowitz KA, Brown AE, Telzak EE, Gottlieb S, Armstrong D. Pneumocystis carinii pneumonia among patients without AIDS at a cancer hospital. *JAMA* 1992; **267**: 832–7.
- 8 Cregan P, Yamamoto A, Lum A, VanDerHeide T, MacDonald M, Pulliam L. Comparison of four methods for rapid detection of Pneumocystis carinii in respiratory specimens. *J Clin Microbiol* 1990; **28**: 2432–6.
- 9 Limper AH, Offord KP, Smith TF, Martin WJ 2nd. Pneumocystis carinii pneumonia. Differences in lung parasite number and inflammation in patients with and without AIDS. *Am Rev Respir Dis* 1989; **140**: 1204–9.
- 10 Robberts FJ, Liebowitz LD, Chalkley LJ. Polymerase chain reaction detection of Pneumocystis jirovecii: evaluation of 9 assays. *Diagn Microbiol Infect Dis* 2007; **58**: 385–92.
- 11 Lu Y, Ling G, Qiang C, Ming Q, Wu C, Wang K *et al*. PCR diagnosis of Pneumocystis pneumonia: a bivariate meta-analysis. *J Clin Microbiol* 2011; **49**: 4361–3.
- 12 Fan LC, Lu HW, Cheng KB, Li HP, Xu JF. Evaluation of PCR in bronchoalveolar lavage fluid for diagnosis of Pneumocystis jirovecii pneumonia: a bivariate meta-analysis and systematic review. *PLoS ONE* 2013; **8**: e73099.
- 13 Huggett JF, Taylor MS, Kocjan G, Evans HE, Morris-Jones S, Gant V *et al*. Development and evaluation of a real-time PCR assay for detection of Pneumocystis jirovecii DNA in bronchoalveolar lavage fluid of HIV-infected patients. *Thorax* 2008; **63**: 154–9.
- 14 Matsumura Y, Ito Y, Iinuma Y, Yasuma K, Yamamoto M, Matsushima A *et al*. Quantitative real-time PCR and the (1- > 3)-beta-D-glucan assay for differentiation between Pneumocystis jirovecii pneumonia and colonization. *Clin Microbiol Infect* 2012; **18**: 591–7.
- 15 Botterel F, Cabaret O, Foulet F, Cordonnier C, Costa JM, Bretagne S. Clinical significance of quantifying Pneumocystis jirovecii DNA by using real-time PCR in bronchoalveolar lavage fluid from immunocompromised patients. *J Clin Microbiol* 2012; **50**: 227–31.
- 16 Muhlethaler K, Bogli-Stubler K, Wasmer S, von Garnier C, Dumont P, Rauch A *et al*. Quantitative PCR to diagnose Pneumocystis pneumonia in immunocompromised non-HIV patients. *Eur Respir J* 2012; **39**: 971–8.
- 17 Alanio A, Desoubeaux G, Sarfati C, Hãmene S, Bergeron A, Azoulay E *et al*. Real-time PCR assay-based strategy for differentiation between active Pneumocystis jirovecii pneumonia and colonization in immunocompromised patients. *Clin Microbiol Infect* 2011; **17**: 1531–7.
- 18 Maillet M, Maubon D, Brion JP, Francois P, Molina L, Stahl JP *et al*. Pneumocystis jirovecii (Pj) quantitative PCR to differentiate Pj pneumonia from Pj colonization in immunocompromised patients. *Eur J Clin Microbiol Infect Dis* 2013.
- 19 Damiani C, Le Gal S, Da Costa C, Virmaux M, Nevez G, Totet A. Combined quantification of pulmonary Pneumocystis jirovecii DNA and serum (1- > 3)-beta-D-glucan for differential diagnosis of pneumocystis pneumonia and Pneumocystis colonization. *J Clin Microbiol* 2013; **51**: 3380–8.
- 20 Teh BW, Azzato FA, Lingaratnam SM, Thursky KA, Slavin MA, Worth LJ. Molecular diagnosis of Pneumocystis jirovecii in patients with malignancy: clinical significance of quantitative polymerase chain reaction. *Med Mycol* 2014; **52**(4): 427–432.
- 21 Larsen HH, Masur H, Kovacs JA, Gill VJ, Silcott VA, Kogulan P *et al*. Development and evaluation of a quantitative, touch-down, real-time PCR assay for diagnosing Pneumocystis carinii pneumonia. *J Clin Microbiol* 2002; **40**: 490–4.
- 22 Vogel MN, Vatlach M, Weissgerber P, Goepfert B, Claussen CD, Hetzel J *et al*. HRCT-features of Pneumocystis jirovecii pneumonia and their evolution before and after treatment in non-HIV immunocompromised patients. *Eur J Radiol* 2012; **81**: 1315–20.
- 23 Kanne JP, Yandow DR, Meyer CA. Pneumocystis jirovecii pneumonia: high-resolution CT findings in patients with and without HIV infection. *AJR Am J Roentgenol* 2012; **198**: W555–61.
- 24 Tasaka S, Tokuda H, Sakai F, Fujii T, Tateda K, Johkoh T *et al*. Comparison of clinical and radiological features of pneumocystis pneumonia between malignancy cases and acquired immunodeficiency syndrome cases: a multicenter study. *Intern Med* 2010; **49**: 273–81.
- 25 Fujii T, Nakamura T, Iwamoto A. Pneumocystis pneumonia in patients with HIV infection: clinical manifestations, laboratory findings, and radiological features. *J Infect Chemother* 2007; **13**: 1–7.
- 26 Reid AB, Chen SC, Worth LJ. Pneumocystis jirovecii pneumonia in non-HIV-infected patients: new risks and diagnostic tools. *Curr Opin Infect Dis* 2011; **24**: 534–44.
- 27 Haeusler GM, Slavin MA, Seymour JF, Lingaratnam S, Teh BW, Tam CS *et al*. Late-onset Pneumocystis jirovecii pneumonia post-fludarabine, cyclophosphamide and rituximab: implications for prophylaxis. *Eur J Haematol* 2013; **91**: 157–63.
- 28 Nakazato T, Mihara A, Sanada Y, Suzuki K, Aisa Y, Iwabuchi M *et al*.

- Pneumocystis jiroveci pneumonia detected by FDG-PET. *Ann Hematol* 2010; **89**: 839–40.
- 29 Hughes WT, Kuhn S, Chaudhary S, Feldman S, Verzosa M, Aur RJ *et al*. Successful chemoprophylaxis for Pneumocystis carinii pneumonitis. *N Engl J Med* 1977; **297**: 1419–26.
- 30 Green H, Paul M, Vidal L, Leibovici L. Prophylaxis of Pneumocystis pneumonia in immunocompromised non-HIV-infected patients: systematic review and meta-analysis of randomized controlled trials. *Mayo Clin Proc* 2007; **82**: 1052–9.
- 31 Ross LE, Hall IJ African American primary care physicians' prostate cancer screening practices. *J Prim Care Community Health* 2014; **5**: 36–43.
- 32 Neumann S, Krause SW, Maschmeyer G, Schiel X, von Lilienfeld-Toal M, Infectious Diseases Working Party (AGIHO) *et al*. Primary prophylaxis of bacterial infections and Pneumocystis jirovecii pneumonia in patients with hematological malignancies and solid tumors: guidelines of the Infectious Diseases Working Party (AGIHO) of the German Society of Hematology and Oncology (DGHO). *Ann Hematol* 2013; **92**: 433–42.
- 33 Peters SG, Prakash UB. Pneumocystis carinii pneumonia. Review of 53 cases. *Am J Med* 1987; **82**: 73–8.
- 34 Arend SM, Kroon FP, van't Wout JW. Pneumocystis carinii pneumonia in patients without AIDS, 1980 through 1993. An analysis of 78 cases. *Arch Intern Med* 1995; **155**: 2436–41.
- 35 Wondergem MJ, Grunberg K, Wittgen BP, Sonneveld P, Zweegman S. Interstitial pneumonitis caused by Pneumocystis jirovecii pneumonia (PCP) during bortezomib treatment. *Histopathology* 2009; **54**: 631–3.
- 36 Shimoma J, Yanagisawa K, Saito B, Kawakami K, Nakamaki T, Tomoyasu S. A case of IgG kappa-type multiple myeloma complicated with carinii pneumonia following thalidomide administration. *Jpn J Antibiot* 2006; **59**: 410–12.
- 37 McKenzie RS, Glenn LD, Goldsmith JC. Pneumocystis carinii pneumonia complicating multiple myeloma. *Chest* 1991; **99**: 656–9.
- 38 Hardak E, Oren I, Dann EJ, Yigla M, Faibish T, Rowe JM *et al*. The increased risk for pneumocystis pneumonia in patients receiving rituximab-CHOP-14 can be prevented by the administration of trimethoprim/sulfamethoxazole: a single-center experience. *Acta Haematol* 2012; **127**: 110–14.
- 39 Chang H, Yeh HC, Su YC, Lee MH. Pneumocystis jiroveci pneumonia in patients with non-Hodgkin's lymphoma receiving chemotherapy containing rituximab. *J Chin Med Assoc* 2008; **71**: 579–82.
- 40 Venhuizen AC, Hustinx WN, van Houte AJ, Veth G, van der Griend R. Three cases of Pneumocystis jirovecii pneumonia (PCP) during first-line treatment with rituximab in combination with CHOP-14 for aggressive B-cell non-Hodgkin's lymphoma. *Eur J Haematol* 2008; **80**: 275–6.
- 41 Huang YC, Liu CJ, Liu CY, Pai JT, Hong YC, Teng HW *et al*. Low absolute lymphocyte count and addition of rituximab confer high risk for interstitial pneumonia in patients with diffuse large B-cell lymphoma. *Ann Hematol* 2011; **90**: 1145–51.
- 42 Kamel S, O'Connor S, Lee N, Filshie R, Nandurkar H, Tam CS. High incidence of Pneumocystis jirovecii pneumonia in patients receiving biweekly rituximab and cyclophosphamide, adriamycin, vincristine, and prednisone. *Leuk Lymphoma* 2010; **51**: 797–801.
- 43 Kolstad A, Holte H, Fossa A, Lauritzen GF, Gaustad P, Torfoss D. Pneumocystis jirovecii pneumonia in B-cell lymphoma patients treated with the rituximab-CHOEP-14 regimen. *Haematologica* 2007; **92**: 139–40.
- 44 Lim KH, Yoon HI, Kang YA, Lee KW, Kim JH, Bang SM *et al*. Severe pulmonary adverse effects in lymphoma patients treated with cyclophosphamide, doxorubicin, vincristine, and prednisone (CHOP) regimen plus rituximab. *Korean J Intern Med* 2010; **25**: 86–92.
- 45 Kim T, Choi SH, Kim SH, Jeong JY, Woo JH, Kim YS *et al*. Point prevalence of Pneumocystis pneumonia in patients with non-Hodgkin lymphoma according to the number of cycles of R-CHOP chemotherapy. *Ann Hematol* 2013; **92**: 231–8.
- 46 Ennishi D, Terui Y, Yokoyama M, Mishima Y, Takahashi S, Takeuchi K *et al*. Increased incidence of interstitial pneumonia by CHOP combined with rituximab. *Int J Hematol* 2008; **87**: 393–7.
- 47 Garcia-Noblejas A, Velasco A, Garcia-Leon N, Cananta-Ortiz J, Steegmann JL. Pneumocystis jirovecii pneumonia as first manifestation of late relapse angioimmunoblastic T-cell lymphoma. *Leuk Res* 2011; **35**: e143–4.
- 48 Hughes WT, Feldman S, Aur RJ, Verzosa MS, Hustu HO, Simone JV. Intensity of immunosuppressive therapy and the incidence of Pneumocystis carinii pneumonitis. *Cancer* 1975; **36**: 2004–9.
- 49 De Castro N, Neuville S, Sarfati C, Ribaud P, Derouin F, Gluckman E *et al*. Occurrence of Pneumocystis jirovecii pneumonia after allogeneic stem cell transplantation: a 6-year retrospective study. *Bone Marrow Transplant* 2005; **36**: 879–83.
- 50 Meyers JD, Flournoy N, Thomas ED. Nonbacterial pneumonia after allogeneic marrow transplantation: a review of ten years' experience. *Rev Infect Dis* 1982; **4**: 1119–32.
- 51 Yale SH, Limper AH. Pneumocystis carinii pneumonia in patients without acquired immunodeficiency syndrome: associated illness and prior corticosteroid therapy. *Mayo Clin Proc* 1996; **71**: 5–13.
- 52 Thursky KA, Worth LJ, Seymour JF, Miles Prince H, Slavin MA. Spectrum of infection, risk and recommendations for prophylaxis and screening among patients with lymphoproliferative disorders treated with alemtuzumab. *Br J Haematol* 2006; **132**: 3–12.
- 53 Tadmor T, McLaughlin P, Polliack A. A resurgence of Pneumocystis in aggressive lymphoma treated with R-CHOP-14: the price of a dose-dense regimen? *Leuk Lymphoma* 2010; **51**: 737–8.
- 54 Kalin M, Kristinsson SY, Cherif H, Lebbad M, Bjorkholm M. Fatal pneumocystis jiroveci pneumonia in ABVD-treated Hodgkin lymphoma patients. *Ann Hematol* 2010; **89**: 523–5.
- 55 Lingaratnam SM, Slavin MA, Thursky KA, Teh BW, Haeusler GM, Seymour JF *et al*. Pneumocystis jirovecii pneumonia associated with gemcitabine chemotherapy: experience at an Australian centre and recommendations for targeted prophylaxis. *Leuk Lymphoma* 2014; Epub ahead of print.
- 56 Hashimoto K, Kobayashi Y, Asakura Y, Mori M, Azuma T, Maruyama D *et al*. Pneumocystis jirovecii pneumonia in

- relation to CD4 + lymphocyte count in patients with B-cell non-Hodgkin lymphoma treated with chemotherapy. *Leuk Lymphoma* 2010; **51**: 1816–21.
- 57 Seymour JF. Concerns regarding data supporting recent *Pneumocystis jirovecii* prophylaxis guidelines. *Ann Hematol* 2013; **92**: 1569–70.
- 58 Katsuya H, Suzumiya J, Sasaki H, Ishitsuka K, Shibata T, Takamatsu Y *et al.* Addition of rituximab to cyclophosphamide, doxorubicin, vincristine, and prednisolone therapy has a high risk of developing interstitial pneumonia in patients with non-Hodgkin lymphoma. *Leuk Lymphoma* 2009; **50**: 1818–23.
- 59 Worth LJ, Dooley MJ, Seymour JF, Mileschkin L, Slavin MA, Thursky KA. An analysis of the utilisation of chemoprophylaxis against *Pneumocystis jirovecii* pneumonia in patients with malignancy receiving corticosteroid therapy at a cancer hospital. *Br J Cancer* 2005; **92**: 867–72.
- 60 Roblot F, Godet C, Le Moal G, Garo B, Faouzi Souala M, Dary M *et al.* Analysis of underlying diseases and prognosis factors associated with *Pneumocystis carinii* pneumonia in immunocompromised HIV-negative patients. *Eur J Clin Microbiol Infect Dis* 2002; **21**: 523–31.
- 61 Roblot F, Le Moal G, Kauffmann-Lacroix C, Bastides F, Boutoille D, Verdon R *et al.* *Pneumocystis jirovecii* pneumonia in HIV-negative patients: a prospective study with focus on immunosuppressive drugs and markers of immune impairment. *Scand J Infect Dis* 2014; **46**: 210–14.
- 62 Sepkowitz KA. *Pneumocystis carinii* pneumonia among patients with neoplastic disease. *Semin Respir Infect* 1992; **7**: 114–21.
- 63 Tasaka S, Tokuda H. *Pneumocystis jirovecii* pneumonia in non-HIV-infected patients in the era of novel immunosuppressive therapies. *J Infect Chemother* 2012; **18**: 793–806.
- 64 American Academy of Paediatrics. *Red book: 2012 Report of the Committee on Infectious Diseases*. 29th edn. Pickering L, ed. Elk Grove Village, IL: American Academy of Paediatrics; 2012.
- 65 Henson JW, Jalaj JK, Walker RW, Stover DE, Fels AO. *Pneumocystis carinii* pneumonia in patients with primary brain tumors. *Arch Neurol* 1991; **48**: 406–9.
- 66 Slivka A, Wen PY, Shea WM, Loeffler JS. *Pneumocystis carinii* pneumonia during steroid taper in patients with primary brain tumors. *Am J Med* 1993; **94**: 216–19.
- 67 Brandes AA, Tosoni A, Cavallo G, Bertorelle R, Gioia V, Franceschi E *et al.* Temozolomide 3 weeks on and 1 week off as first-line therapy for recurrent glioblastoma: phase II study from gruppo italiano cooperativo di neuro-oncologia (GICNO). *Br J Cancer* 2006; **95**: 1155–60.
- 68 Balana C, Lopez-Pousa A, Berrocal A, Yaya-Tur R, Herrero A, Garcia JL *et al.* Phase II study of temozolomide and cisplatin as primary treatment prior to radiotherapy in newly diagnosed glioblastoma multiforme patients with measurable disease. A study of the Spanish Medical Neuro-Oncology Group (GENOM). *J Neurooncol* 2004; **70**: 359–69.
- 69 Stupp R, Dietrich PY, Ostermann Kraljevic S, Pica A, Maillard I, Maeder P *et al.* Promising survival for patients with newly diagnosed glioblastoma multiforme treated with concomitant radiation plus temozolomide followed by adjuvant temozolomide. *J Clin Oncol* 2002; **20**: 1375–82.
- 70 De Vos FY, Gijtenbeek JM, Bleeker-Rovers CP, van Herpen CM. *Pneumocystis jirovecii* pneumonia prophylaxis during temozolomide treatment for high-grade gliomas. *Crit Rev Oncol Hematol* 2013; **85**: 373–82.
- 71 Kim SY, Dabb AA, Glenn DJ, Snyder KM, Chuk MK, Loeb DM. Intravenous pentamidine is effective as second line *Pneumocystis pneumonia* prophylaxis in pediatric oncology patients. *Pediatr Blood Cancer* 2008; **50**: 779–83.
- 72 Sibanda EL, Weller IV, Hakim JG, Cowan FM. Does trimethoprim-sulfamethoxazole prophylaxis for HIV induce bacterial resistance to other antibiotic classes? Results of a systematic review. *Clin Infect Dis* 2011; **52**: 1184–94.
- 73 Rossi MR, Banfi P, Cappuccilli M, Conter V, de Poli D, Piacentini G *et al.* Prospective randomized comparison of two prophylactic regimens with trimethoprim-sulfamethoxazole in leukemic children: a two year study. *Eur J Cancer Clin Oncol* 1987; **23**: 1679–82.
- 74 Caselli D, Petris MG, Rondelli R, Carraro F, Colombini A, Muggeo P *et al.* Single-day trimethoprim/sulfamethoxazole prophylaxis for pneumocystis pneumonia in children with cancer. *J Pediatr* 2013; **164**(2): 389–92.e1
- 75 Hughes WT, Rivera GK, Schell MJ, Thornton D, Lott L. Successful intermittent chemoprophylaxis for *Pneumocystis carinii* pneumonitis. *N Engl J Med* 1987; **316**: 1627–32.
- 76 Muto T, Takeuchi M, Kawaguchi T, Tanaka S, Tsukamoto S, Sakai S *et al.* Low-dose trimethoprim-sulfamethoxazole for *Pneumocystis jirovecii* pneumonia prophylaxis after allogeneic hematopoietic SCT. *Bone Marrow Transplant* 2011; **46**: 1573–5.
- 77 Souza JP, Boeckh M, Gooley TA, Flowers ME, Crawford SW. High rates of *Pneumocystis carinii* pneumonia in allogeneic blood and marrow transplant recipients receiving dapsone prophylaxis. *Clin Infect Dis* 1999; **29**: 1467–71.
- 78 Vasconcelles MJ, Bernardo MV, King C, Weller EA, Antin JH. Aerosolized pentamidine as pneumocystis prophylaxis after bone marrow transplantation is inferior to other regimens and is associated with decreased survival and an increased risk of other infections. *Biol Blood Marrow Transplant* 2000; **6**: 35–43.
- 79 Imrie KR, Prince HM, Couture F, Brandwein JM, Keating A. Effect of antimicrobial prophylaxis on hematopoietic recovery following autologous bone marrow transplantation: ciprofloxacin versus co-trimoxazole. *Bone Marrow Transplant* 1995; **15**: 267–70.
- 80 Fontanet A, Chalandon Y, Roosnek E, Mohty B, Passweg JR. Cotrimoxazole myelotoxicity in hematopoietic SCT recipients: time for reappraisal. *Bone Marrow Transplant* 2011; **46**: 1272–3.
- 81 Relling MV, Fairclough D, Ayers D, Crom WR, Rodman JH, Pui CH *et al.* Patient characteristics associated with high-risk methotrexate concentrations and toxicity. *J Clin Oncol* 1994; **12**: 1667–72.
- 82 Lane BR, Ast JC, Hossler PA, Mindell DP, Bartlett MS, Smith JW *et al.* Dihydropteroate synthase

- polymorphisms in *Pneumocystis carinii*. *J Infect Dis* 1997; **175**: 482–5.
- 83 Dini L, du Plessis M, Freaun J, Fernandez V. High prevalence of dihydropteroate synthase mutations in *Pneumocystis jirovecii* isolated from patients with *Pneumocystis pneumonia* in South Africa. *J Clin Microbiol* 2010; **48**: 2016–21.
- 84 Helweg-Larsen J, Benfield TL, Eugen-Olsen J, Lundgren JD, Lundgren B. Effects of mutations in *Pneumocystis carinii* dihydropteroate synthase gene on outcome of AIDS-associated *P. carinii pneumonia*. *Lancet* 1999; **354**: 1347–51.
- 85 Kazanjian P, Armstrong W, Hossler PA, Burman W, Richardson J, Lee CH et al. *Pneumocystis carinii* mutations are associated with duration of sulfa or sulfone prophylaxis exposure in AIDS patients. *J Infect Dis* 2000; **182**: 551–7.
- 86 Navin TR, Beard CB, Huang L, del Rio C, Lee S, Pieniazek NJ et al. Effect of mutations in *Pneumocystis carinii* dihydropteroate synthase gene on outcome of *P. carinii pneumonia* in patients with HIV-1: a prospective study. *Lancet* 2001; **358**: 545–9.
- 87 Voeller D, Kovacs J, Andrawis V, Chu E, Masur H, Allegra C. Interaction of *Pneumocystis carinii* dihydropteroate synthase with sulfonamides and diaminodiphenyl sulfone (dapson). *J Infect Dis* 1994; **169**: 456–9.
- 88 Cruciani M, Gatti G, Mengoli C, Cazzadori A, Lazzarini L, Miletich F et al. Penetration of dapsone into pulmonary lining fluid of human immunodeficiency virus type 1-infected patients. *Antimicrob Agents Chemother* 1997; **41**: 1077–81.
- 89 Hughes WT. Use of dapsone in the prevention and treatment of *Pneumocystis carinii pneumonia*: a review. *Clin Infect Dis* 1998; **27**: 191–204.
- 90 Slavin MA, Hoy JF, Stewart K, Pettinger MB, Lucas CR, Kent SJ. Oral dapsone versus nebulized pentamidine for *Pneumocystis carinii pneumonia* prophylaxis: an open randomized prospective trial to assess efficacy and haematological toxicity. *AIDS* 1992; **6**: 1169–74.
- 91 Green H, Paul M, Vidal L, Leibovici L. Prophylaxis for *Pneumocystis pneumonia* (PCP) in non-HIV immunocompromised patients. *Cochrane Database Syst Rev* 2007; (3): CD 005590.
- 92 DeMasi JM, Cox JA, Leonard D, Koh AY, Aquino VM. Intravenous pentamidine is safe and effective as primary *pneumocystis pneumonia* prophylaxis in children and adolescents undergoing hematopoietic stem cell transplantation. *Pediatr Infect Dis J* 2013; **32**: 933–6.
- 93 Colby C, McAfee S, Sackstein R, Finkelstein D, Fishman J, Spitzer T. A prospective randomized trial comparing the toxicity and safety of atovaquone with trimethoprim/sulfamethoxazole as *Pneumocystis carinii pneumonia* prophylaxis following autologous peripheral blood stem cell transplantation. *Bone Marrow Transplant* 1999; **24**: 897–902.
- 94 *Therapeutic Guidelines: Antibiotic, Version 15*. Melbourne, Australia: Therapeutic Guidelines Limited; 2014.
- 95 Safrin S, Finkelstein DM, Feinberg J, Frame P, Simpson G, Wu A et al. Comparison of three regimens for treatment of mild to moderate *Pneumocystis carinii pneumonia* in patients with AIDS. A double-blind, randomized, trial of oral trimethoprim-sulfamethoxazole, dapsone-trimethoprim, and clindamycin-primaquine. ACTG 108 Study Group. *Ann Intern Med* 1996; **124**: 792–802.
- 96 Hughes W, Leoung G, Kramer F, Bozzette SA, Safrin S, Frame P et al. Comparison of atovaquone (566C80) with trimethoprim-sulfamethoxazole to treat *Pneumocystis carinii pneumonia* in patients with AIDS. *N Engl J Med* 1993; **328**: 1521–7.
- 97 Masur H. Prevention and treatment of *pneumocystis pneumonia*. *N Engl J Med* 1992; **327**: 1853–60.
- 98 Corticosteroid therapy for AIDS-associated PCP. *Am Fam Physician* 1990; **42**: 1672–5.
- 99 Bozzette SA, Sattler FR, Chiu J, Wu AW, Gluckstein D, Kemper C et al. A controlled trial of early adjunctive treatment with corticosteroids for *Pneumocystis carinii pneumonia* in the acquired immunodeficiency syndrome. California Collaborative Treatment Group. *N Engl J Med* 1990; **323**: 1451–7.
- 100 Gagnon S, Boota AM, Fischl MA, Baier H, Kirksey OW, La Voie L. Corticosteroids as adjunctive therapy for severe *Pneumocystis carinii pneumonia* in the acquired immunodeficiency syndrome. A double-blind, placebo-controlled trial. *N Engl J Med* 1990; **323**: 1444–50.
- 101 Montaner JS, Lawson LM, Levitt N, Belzberg A, Schechter MT, Ruedy J. Corticosteroids prevent early deterioration in patients with moderately severe *Pneumocystis carinii pneumonia* and the acquired immunodeficiency syndrome (AIDS). *Ann Intern Med* 1990; **113**: 14–20.
- 102 el Sadr W, Simberloff MS. Survival and prognostic factors in severe *Pneumocystis carinii pneumonia* requiring mechanical ventilation. *Am Rev Respir Dis* 1988; **137**: 1264–7.
- 103 Moon SM, Kim T, Sung H, Kim MN, Kim SH, Choi SH et al. Outcomes of moderate-to-severe *Pneumocystis pneumonia* treated with adjunctive steroid in non-HIV-infected patients. *Antimicrob Agents Chemother* 2011; **55**: 4613–18.
- 104 Delclaux C, Zahar JR, Amraoui G, Leleu G, Lebargy F, Brochard L et al. Corticosteroids as adjunctive therapy for severe *Pneumocystis carinii pneumonia* in non-human immunodeficiency virus-infected patients: retrospective study of 31 patients. *Clin Infect Dis* 1999; **29**: 670–2.
- 105 Pareja JG, Garland R, Koziel H. Use of adjunctive corticosteroids in severe adult non-HIV *Pneumocystis carinii pneumonia*. *Chest* 1998; **113**: 1215–24.
- 106 Klein NC, Duncanson FP, Lenox TH, Forszpaniak C, Sherer CB, Quentzel H et al. Trimethoprim-sulfamethoxazole versus pentamidine for *Pneumocystis carinii pneumonia* in AIDS patients: results of a large prospective randomized treatment trial. *AIDS* 1992; **6**: 301–5.
- 107 Sattler FR, Cowan R, Nielsen DM, Ruskin J. Trimethoprim-sulfamethoxazole compared with pentamidine for treatment of *Pneumocystis carinii pneumonia* in the acquired immunodeficiency syndrome. A prospective, noncrossover study. *Ann Intern Med* 1988; **109**: 280–7.
- 108 Helweg-Larsen J, Benfield T, Atzori C, Miller RF. Clinical efficacy of first- and second-line treatments for HIV-associated *Pneumocystis jirovecii pneumonia*: a tri-centre cohort study.

- J Antimicrob Chemother* 2009; **64**: 1282–90.
- 109 Conte JE Jr, Chernoff D, Feigal DW Jr, Joseph P, McDonald C, Golden JA. Intravenous or inhaled pentamidine for treating *Pneumocystis carinii* pneumonia in AIDS. A randomized trial. *Ann Intern Med* 1990; **113**: 203–9.
- 110 Toma E, Thorne A, Singer J, Raboud J, Lemieux C, Trottier S *et al*. Clindamycin with primaquine vs. Trimethoprim-sulfamethoxazole therapy for mild and moderately severe *Pneumocystis carinii* pneumonia in patients with AIDS: a multicenter, double-blind, randomized trial (CTN 004). CTN-PCP Study Group. *Clin Infect Dis* 1998; **27**: 524–30.
- 111 Medina I, Mills J, Leoung G, Hopewell PC, Lee B, Modin G *et al*. Oral therapy for *Pneumocystis carinii* pneumonia in the acquired immunodeficiency syndrome. A controlled trial of trimethoprim-sulfamethoxazole versus trimethoprim-dapsone. *N Engl J Med* 1990; **323**: 776–82.
- 112 Kottom TJ, Limper AH. Cell wall assembly by *Pneumocystis carinii*. Evidence for a unique gsc-1 subunit mediating beta -1,3-glucan deposition. *J Biol Chem* 2000; **275**: 40628–34.
- 113 Mu XD, Que CL, He B, Wang GF, Li HC. Caspofungin in salvage treatment of severe pneumocystis pneumonia: case report and literature review. *Chin Med J (Engl)* 2009; **122**: 996–9.
- 114 Kim T, Hong HL, Lee YM, Sung H, Kim SH, Choi SH *et al*. Is caspofungin really an effective treatment for *Pneumocystis jirovecii* pneumonia in immunocompromised patients without human immunodeficiency virus infection? Experiences at a single center and a literature review. *Scand J Infect Dis* 2013; **45**: 484–8.
- 115 Kamboj M, Weinstock D, Sepkowitz KA. Progression of *Pneumocystis jirovecii* pneumonia in patients receiving echinocandin therapy. *Clin Infect Dis* 2006; **43**: e92–4.
- 116 Beltz K, Kramm CM, Laws HJ, Schrotten H, Wessalowski R, Gobel U. Combined trimethoprim and caspofungin treatment for severe *Pneumocystis jirovecii* pneumonia in a five year old boy with acute lymphoblastic leukemia. *Klin Padiatr* 2006; **218**: 177–9.
- 117 Ceballos ME, Ortega M, Andresen M, Wozniak A, Garcia P, Balcells ME. Successful treatment with echinocandin in an HIV-infected individual failing first-line therapy for *Pneumocystis jirovecii* pneumonia. *AIDS* 2011; **25**: 2192–3.
- 118 Schmoldt S, Schuegger R, Wendler T, Huber I, Sollner H, Hogardt M *et al*. Molecular evidence of nosocomial *Pneumocystis jirovecii* transmission among 16 patients after kidney transplantation. *J Clin Microbiol* 2008; **46**: 966–71.
- 119 Phipps LM, Chen SC, Kable K, Halliday CL, Firacative C, Meyer W *et al*. Nosocomial *Pneumocystis jirovecii* pneumonia: lessons from a cluster in kidney transplant recipients. *Transplantation* 2011; **92**: 1327–34.
- 120 de Boer MG, de Fijter JW, Kroon FP. Outbreaks and clustering of *Pneumocystis* pneumonia in kidney transplant recipients: a systematic review. *Med Mycol* 2011; **49**: 673–80.
- 121 Bensousan T, Garo B, Islam S, Bourbigot B, Cledes J, Garre M. Possible transfer of *Pneumocystis carinii* between kidney transplant recipients. *Lancet* 1990; **336**: 1066–7.
- 122 Damiani C, Choukri F, Le Gal S, Menotti J, Sarfati C, Nevez G *et al*. Possible nosocomial transmission of *Pneumocystis jirovecii*. *Emerg Infect Dis* 2012; **18**: 877–8.
- 123 Choukri F, Menotti J, Sarfati C, Lucet JC, Nevez G, Garin YJ *et al*. Quantification and spread of *Pneumocystis jirovecii* in the surrounding air of patients with *Pneumocystis* pneumonia. *Clin Infect Dis* 2010; **51**: 259–65.
- 124 Bartlett MS, Vermund SH, Jacobs R, Durant PJ, Shaw MM, Smith JW *et al*. Detection of *Pneumocystis carinii* DNA in air samples: likely environmental risk to susceptible persons. *J Clin Microbiol* 1997; **35**: 2511–13.
- 125 Le Gal S, Damiani C, Rouille A, Grall A, Treguer L, Virmaux M *et al*. A cluster of *Pneumocystis* infections among renal transplant recipients: molecular evidence of colonized patients as potential infectious sources of *Pneumocystis jirovecii*. *Clin Infect Dis* 2012; **54**: e62–71.
- 126 Vargas SL, Ponce CA, Gigliotti F, Ulloa AV, Prieto S, Munoz MP *et al*. Transmission of *Pneumocystis carinii* DNA from a patient with *P. carinii* pneumonia to immunocompetent contact health care workers. *J Clin Microbiol* 2000; **38**: 1536–8.
- 127 Medrano FJ, Montes-Cano M, Conde M, de la Horra C, Respaldiza N, Gasch A *et al*. *Pneumocystis jirovecii* in general population. *Emerg Infect Dis* 2005; **11**: 245–50.
- 128 Chabe M, Dei-Cas E, Creusy C, Fleurisse L, Respaldiza N, Camus D *et al*. Immunocompetent hosts as a reservoir of pneumocystis organisms: histological and rt-PCR data demonstrate active replication. *Eur J Clin Microbiol Infect Dis* 2004; **23**: 89–97.
- 129 Rostved AA, Sassi M, Kurtzhals JA, Sorensen SS, Rasmussen A, Ross C *et al*. Outbreak of pneumocystis pneumonia in renal and liver transplant patients caused by genotypically distinct strains of *Pneumocystis jirovecii*. *Transplantation* 2013; **96**: 834–42.
- 130 Gianella S, Haeberli L, Joos B, Ledergerber B, Wuthrich RP, Weber R *et al*. Molecular evidence of interhuman transmission in an outbreak of *Pneumocystis jirovecii* pneumonia among renal transplant recipients. *Transpl Infect Dis* 2010; **12**: 1–10.
- 131 Nankivell BJ, Firacative C, Kable K, Chen SC, Meyer W. Molecular epidemiology linking multihospital clusters of opportunistic *Pneumocystis jirovecii* pneumonia. *Clin Infect Dis* 2013; **57**: 1058–9.